

Absorption of neutral and basic amino acids across the body surface of two annelid species

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KURZFASSUNG: Absorption von neutralen und basischen Aminosäuren durch die Körperoberfläche zweier Annelidenarten. In Versuchen zur Aufnahme verschiedener Aminosäuren durch die Körperoberfläche bei dem Oligochaeten *Enchytraeus albidus* und dem Polychaeten *Nereis diversicolor* betrug die Absorptionsrate von ^{14}C -L-Arginin aus einer Umgebungskonzentration von $10\ \mu\text{M}$ nur wenige Prozent der für Glycin und L-Valin bestimmten Werte. Nach einer halbstündigen Versuchsdauer waren neutrale ^{14}C -Aminosäuren im Organismus 9- bis 15fach gegenüber dem Medium angereichert, die L-Argininverteilung blieb dagegen unterhalb des Äquilibriums. Die konzentrationsabhängige Aufnahmekurve für L-Arginin bestand bei *E. albidus* aus einer Diffusions- und einer Sättigungskomponente, die durch andere Aminosäuren hemmbar war. Die stärkste Inhibition erfolgte durch basische Aminosäuren. Eine erhebliche Hemmung wurde auch durch neutrale Aminosäuren bewirkt. L-Glutamin und L-Asparagin verminderten die L-Argininaufnahme nur geringfügig. Die Gegenwart von sauren, β - und γ -Aminosäuren sowie organischen Säuren und Kohlenhydraten hatte nur einen unbedeutenden oder gar keinen Einfluß zur Folge. Da die L-Argininaufnahme nicht dem Einfluß von Stoffwechsellinhibitoren (NaCN und Ouabain) unterlag und Na^+ -unabhängig war, erfüllt die Sättigungskomponente des L-Arginintransports die Merkmale der "erleichterten Diffusion". Der Unterschied zu neutralen Aminosäuren, welche aktiv transportiert werden, wird diskutiert.

INTRODUCTION

A variety of soft-bodied marine invertebrates utilizes an additional food source from uptake of organic matter dissolved in sea water. This mode of nutrition occurs independently of feeding particulate nutrients and digestion in alimentary canals. From a number of experiments conducted in recent years, it can be concluded that absorption of several amino acids and glucose may proceed from extremely low concentrations against considerable concentration gradients of roughly 4 to 6 orders of magnitude, and that absorbed organic matter is accumulated and metabolically utilized (see STEPHENS, 1972).

Investigations of several marine and brackish water invertebrate species of different phyla have shown that absorption rates of amino acids are positively correlated to ambient salinity levels (STEPHENS, 1964; STEPHENS & VIRKAR, 1966; SHICK, 1973; BAMFORD & CAMPBELL, 1976; SIEBERS & BULNHEIM, in press), implying that fresh

water invertebrates cannot obtain significant nutritional profit from uptake of dissolved organic matter. The mechanism which underlies the salinity dependence of amino acid absorption is probably similar to the sodium dependence of solute transport across animal membranes (SCHULTZ & CURRAN, 1970).

Results from various cross-inhibition experiments with single animal cells and absorbing tissues, concerning specificities of amino acid absorption suggest separate absorbing systems, which recognize the α -amino- α -carboxyl configuration of the molecule and its charge. This means that separate uptake systems exist for neutral, basic, and acidic amino acids. While vertebrate cells and gastrointestinal tissues (WISEMAN, 1974) and the body surface of intestinal parasites (PAPPAS & READ, 1975) have been thoroughly investigated in this respect, appropriate information on the mechanisms operating in the body walls of marine invertebrates is – with few exceptions – still lacking. For the body surface of the oligochaete annelid *Enchytraeus albidus* SIEBERS (in press) demonstrated the existence of an uptake system for neutrally charged α -amino acids, on which basic amino acids have a small, and acidic amino acids, carbohydrates, and organic acids have no influence.

The present paper is devoted to a more detailed analysis of absorption of neutral and basic amino acids across the body surface in the oligochaete *Enchytraeus albidus* and the polychaete *Nereis diversicolor* including the dependence of uptake velocities on energy supply.

MATERIALS AND METHODS

E. albidus HENLE, obtained from earth cultures, was transferred to the bottom filter of all-glass aquaria (15° C, 20 ‰ S) and used for experiments after about two weeks of submersal life. *N. diversicolor* MÜLLER was dug from the surface layer of the bottom of the Sehlendorfer Binnensee, close to the coastline of the western Baltic Sea (Schleswig-Holstein, FRG). Specimens were maintained in a 1 cm high bottom layer of sand in all-glass aquaria (15° C, 20 ‰), and the water was filtered in circuit. Specimens of 3–4 cm body length were taken for experimental use after about 1 week of acclimation to maintaining conditions. Both species were fed with artificial fish food (Tetra Min).

Experimental assays were run in a water volume of 10 ml, containing about 50 mg of worms (about 15–20 individuals of *E. albidus* and 1 individual *N. diversicolor*), 0.2 μ Ci of a uniformly labeled ¹⁴C-amino acid (glycine, L-valine, L-arginine, L-lysine) (Amersham), and 0.1–2.4 μ moles of unlabeled amino acids of highest purity available.

For experiments concerning effects of metabolic inhibitors on amino acid absorption the incubation medium additionally contained 1–10 μ moles of ouabain (G-strophanthin)/10 ml, or 0.5–2.5 μ moles of NaCN/10 ml. When effects of sodium depletion were investigated, the incubation media were 20 ‰ sea water, 348 mM NaCl (the solution corresponds to a salinity of 20 ‰ sea water by aerometry), 87 mM NaCl + 261 mM KCl, 87 mM NaCl + 261 mM LiCl, and 87 mM NaCl + 261 mM choline-Cl. To realize effects of metabolic inhibitors or sodium-depletion, a preincubation period of 4 h (15° C) proved sufficient.

The experimental time for determination of uptake rates was 0.5 h throughout all analyses. The salinity of experimental water (20 ‰ S) and the temperature (15° C) were kept constant.

All investigations were run with 4–7 replicates; differences in uptake rates between varying experimental conditions were statistically analyzed by STUDENT'S t-test.

For determination of uptake rates the experimental incubation medium was removed, worms were rinsed twice with appropriate incubation medium (not containing inactive or active amino acids), dried on filter paper and stored at – 25° C. Counting of absorbed label was performed in a Tracerlab liquid scintillation counter with quench correction by internal standard after solubilizing worms for 1 h at 60° C in soluene-350 (Packard) and addition of 10 ml of counting solution (5 g PPO + 0.3 g POPOP/l of toluol).

Before insertion of worms into experimental incubation media, 200 μ l were removed from 10 ‰ of assays and counted for experimental standard after addition of 200 μ l of distilled water and 10 ml Insta-Gel (Packard).

The experimental water of 20 ‰ S was taken from a 40-l-reservoir, in which it was permanently filtered over gravel at 15° C in the dark and only ultra-filtered. Regarding previously used precautionary handling to avoid unknown zero levels of dissolved amino acids (passing a bed of charcoal, shaking with permutite, centrifugation, and ultra-filtration), this handling proved to be unnecessary, since only ultra-filtration of the permanently filtered water revealed identical experimental results. Sea water which is stored in the dark for a long period contains only traces of dissolved amino acids (BOHLING, 1970, 1972), probably due to bacterial degradation.

RESULTS

Uptake velocities

Concentration dependent L-arginine uptake velocities in *E. albidus* are presented in Figure 1. The uptake curve (A) demonstrates a diffusion component, especially at higher substrate concentrations. After subtraction of the diffusion component from uptake curve (A) a saturable component is obtained (B) with a maximum velocity

Table 1

Enchytraeus albidus and *Nereis diversicolor*. Comparison of absorption capacities from 10 μ M solutions of glycine, L-valine, and L-arginine (15° C, 20 ‰ S, n = 4)

Absorbed amino acid	Absorption rate (n moles/g/h)		Distribution ratio	
	<i>Enchytraeus albidus</i>	<i>Nereis diversicolor</i>	<i>Enchytraeus albidus</i>	<i>Nereis diversicolor</i>
glycine	211 \pm 20	267 \pm 13	10.6	13.4
L-valine	179 \pm 19	298 \pm 40	8.9	14.9
L-arginine	4.8 \pm 0.2	10.9 \pm 2.7	0.24	0.54

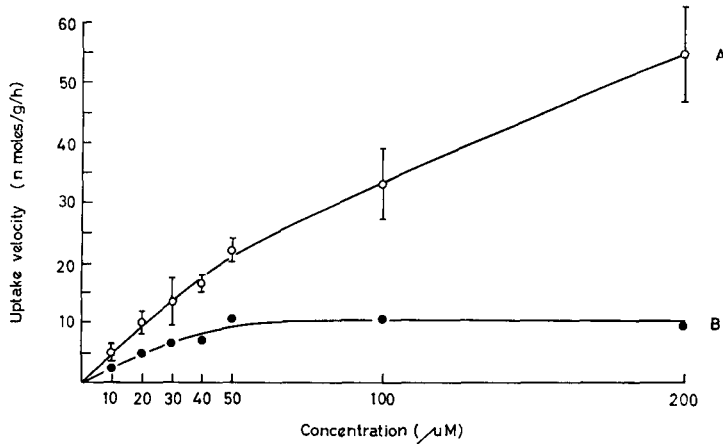


Fig. 1: *E. albidus*. Concentration-dependent uptake velocity of L-arginine (A, \circ) (15°C , 20‰ S , $n = 4$). The lower curve (B, \bullet) demonstrates the saturable component after correction for diffusion

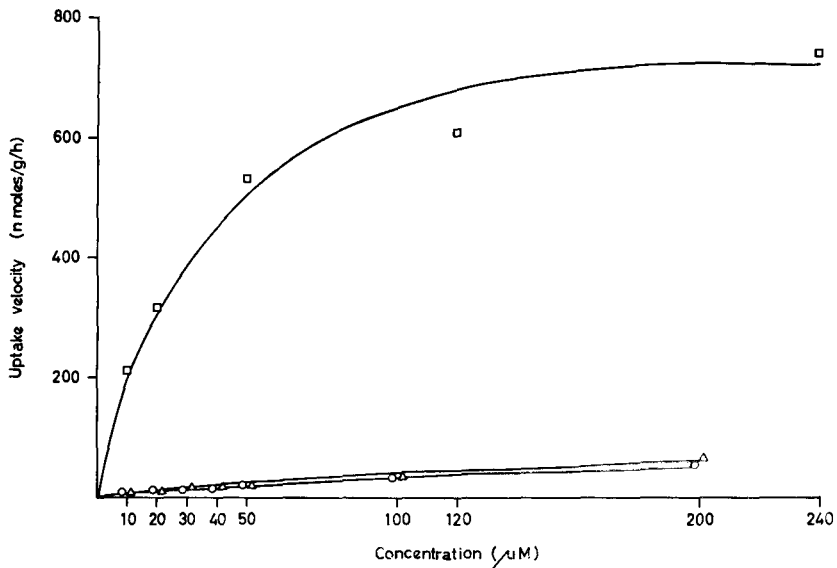


Fig. 2: *E. albidus*. Comparison of concentration-dependent uptake velocities of glycine (\square), L-arginine (\circ), and L-lysine (\triangle) (15°C , 20‰ S , $n = 4$)

(v_{max}) of 10.5 nmoles/g/h and a transport constant of $21\ \mu\text{M}$ (Lineweaver-Burk-plot). Uptake rates of basic amino acids, as exemplified for L-arginine and L-lysine in Figure 2, are much lower than uptake rates of neutral amino acids, as exemplified for glycine. (The uptake curve of L-valine does not differ greatly from that of glycine). At an external concentration of $10\ \mu\text{M}$ (15°C , 20‰ S) arginine uptake amounts to only 2.3% of glycine uptake and 2.7% of L-valine uptake in *E. albidus* (Table 1).

This great difference in uptake rates according to the net charge of the transported amino acid is also valid for the polychaete *Nereis diversicolor* (Table 1). Uptake of the neutral amino acids glycine and L-valine and positively charged L-arginine was investigated under identical experimental conditions in *Enchytraeus* and *Nereis*. In *N. diversicolor* uptake rates are significantly higher by 27 % for glycine, by 66 % for L-valine, and by 127 % for L-arginine than in *E. albidus*. However, this direct comparison is only partly admissible, because the experimental assays contained about 50 mg of fresh weight; they comprised 15–20 individuals of *E. albidus* and only one individual of *N. diversicolor* which is supplied with a large parapodial body surface area. A much lower uptake rate of L-arginine compared to glycine and L-valine is also found in the polychaete.

The distribution ratios (radioactivity/50 mg of worms : radioactivity/50 μ l of ambient water after 0.5 h of experimental time) range between 8.9 and 14.9 for glycine and L-valine and between 0.24 and 0.54 for L-arginine. They demonstrate a concentrating process for uptake of neutral amino acids. L-arginine distribution ratios, however, do not exceed the equilibrium (distribution ratio = 1) (Table 1). Even when assuming an intracellular water amount of roughly 50 % of fresh weight, which contains more than 95 % of total free amino acids – this amino acid distribution seems to be valid for most marine invertebrates (FLORKIN, 1969) – distribution ratios would be roughly doubled, but would not significantly exceed the equilibrium distribution ratio of 1.

The differences in amino acid absorption between *E. albidus* and *N. diversicolor* are significant (Table 1); however, the values measured are of the same order of magnitude. The oligochaete and the polychaete reveal the same low uptake rates of L-arginine, high distribution ratios of neutral amino acids and small distribution ratios

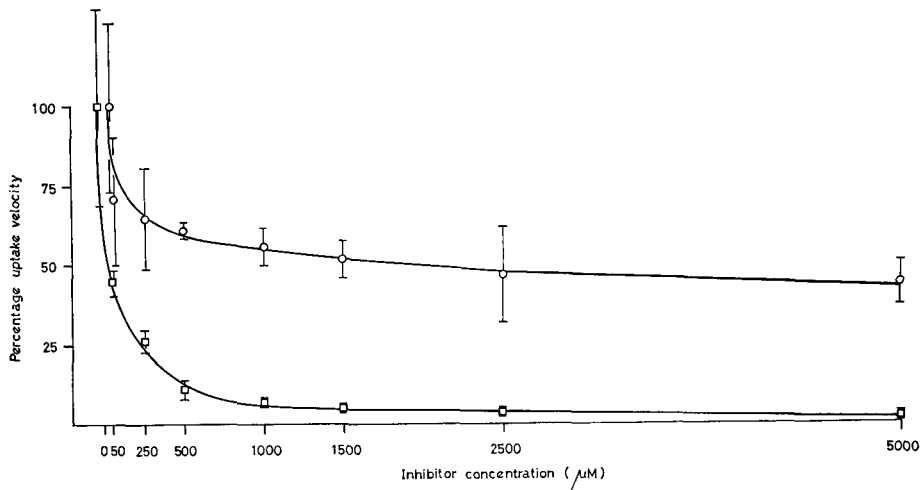


Fig. 3: *E. albidus*. Inhibition of L-arginine uptake by increasing concentrations of L-alanine (○) and inhibition of glycine uptake by increasing concentrations of L-methionine (◻). L-arginine and glycine were absorbed from 50 μ M solutions in 20 ‰ sea water at 15° C, n = 4

of L-arginine, which do not exceed the equilibrium distribution ratio of 1. This correspondence in absorption data of amino acids indicates that the oligochaete *E. albidus* may be regarded as a possible model organism for transport studies of amino acids and other solutes in annelids.

The existence of a simple diffusion component of L-arginine uptake in *E. albidus* (Fig. 1) is confirmed by an experiment, in which L-arginine is absorbed from a constant concentration of 50 μM , with L-alanine concentrations increasing from 0 to 5000 μM (Fig. 3). Alanine does not reduce arginine uptake to zero levels. A distinct proportion of L-arginine absorption is not susceptible to inhibition by L-alanine. This proportion is regarded to be the non-saturable simple diffusion component. Glycine absorption, on the contrary, is characterized by saturation kinetics (Fig. 2) without a diffusion component. Increasing concentrations of L-methionine reduce glycine absorption rates to zero levels (Fig. 3). Glycine and L-methionine are regarded to share the same absorptive mechanism for neutral amino acids (SIEBERS, in press).

Specificity of L-arginine transport

When L-arginine is absorbed by *E. albidus* from 50 μM solutions in the presence of 500 μM concentrations of different charged and uncharged α -amino acids, amino acid amides, β - and γ -amino acids, organic acids, and carbohydrates, arginine transport rates are unchanged in the presence of organic acids and carbohydrates (Fig. 4). Arginine uptake is inhibited inconsiderably by acidic amino acids and β - and γ -amino acids. A low, but significant reduction of arginine transport results from the amino acid amides L-glutamine and L-asparagine, while strong reduction of arginine uptake

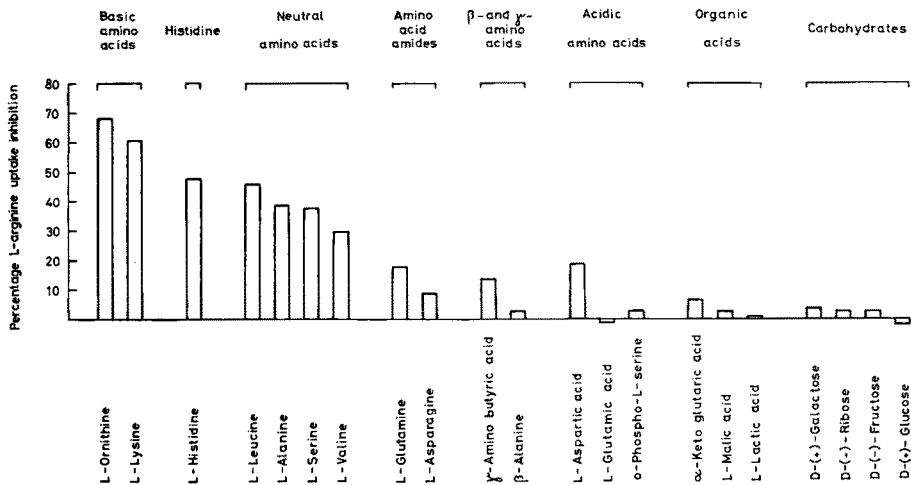


Fig. 4: *E. albidus*. Inhibition of L-arginine uptake from 50 μM solution by 500 μM of charged and uncharged α -amino acids, amino acid amides, β - and γ -amino acids, organic acids, and carbohydrates (15 $^{\circ}$ C, 20‰ S, n = 5)

is observed in the presence of the four neutral amino acids tested (L-leucine, L-alanine, L-serine, and L-valine) and L-histidine. The strongest inhibitions result from the di-basic amino acids L-ornithine and L-lysine.

Effect of metabolic inhibitors and Na-depletion

One of the criteria for the presence of an active transport process is the dependency on energy which is necessary to transport a solute in question against a concentration gradient. A 4-h preincubation of *E. albidus* in 50, 100 and 250 μM NaCN, which is known to act as an inhibitor of respiration, proved sufficient to realize significant reductions in uptake rates of neutral amino acids (glycine and L-valine) (Table 2).

Table 2

Enchytraeus albidus. Effect of metabolic inhibitors (NaCN, Ouabain) on absorption rates of glycine, L-valine, and L-arginine from 10 μM solutions after preincubation in appropriate inhibitor concentrations for 4 h (15° C, 20 % S, n = 7). * indicates statistical significance of differences in uptake rate ($0.05 > p < 0.01$), ** indicates high significance ($p < 0.01$), n.s. indicates no significance ($p > 0.05$) in comparison to absorption rates in the absence of inhibitors

Inhibitor concentrations	Absorption rates, levels of significance					
	Glycine	(%)	L-valine	(%)	L-arginine	(%)
no inhibitor	203 \pm 26	(100)	169 \pm 10	(100)	5.1 \pm 0.4	(100)
50 μM NaCN	138 \pm 18**	(68)	103 \pm 3**	(61)	4.6 \pm 0.5 n.s.	(90)
100 μM NaCN	77 \pm 16**	(38)	71 \pm 7**	(42)	3.7 \pm 0.7**	(73)
250 μM NaCN	45 \pm 14**	(22)	29 \pm 5**	(17)	3.5 \pm 0.7**	(69)
100 μM Ouabain	197 \pm 24 n.s.	(97)	110 \pm 8**	(65)	5.6 \pm 0.5 n.s.	(110)
500 μM Ouabain	142 \pm 26**	(70)	74 \pm 10**	(44)	4.6 \pm 0.6 n.s.	(90)
1000 μM Ouabain	85 \pm 20**	(42)	74 \pm 7**	(44)	4.6 \pm 0.7 n.s.	(90)

At a concentration of 250 μM NaCN, only 22 % of glycine and 17 % of L-valine were absorbed from 10 μM solutions as compared to the controls with no inhibitor present. These results indicate an energy-depending step in the transport of neutral amino acids. Inhibition of glycine and L-valine transport by ouabain (Table 2) demonstrates secondary active transport of neutral amino acids. Ouabain is known as a potent inhibitor of the monovalent cation pump. Inhibition of neutral amino-acid transport by ouabain demonstrates dependency on Na-transport as the energy-supplying step. L-arginine uptake rates, however, are reduced only in a much smaller degree by increasing concentrations of NaCN and ouabain. It must be argued that these substances may have complex poisonous effects, following the inhibitory actions mentioned above, and finally cause the death of the animals. Considering these aspects it seems likely that, in contrast to neutral amino acids, the absorptive step of L-arginine is energy-independent.

In all cases, where active transport of an amino acid or a sugar in an animal tissue has been demonstrated, also Na-dependence was found (SAIER & STILES, 1975). L-arginine uptake in *E. albidus*, however, does not depend on external Na⁺ (Table 3).

Table 3

Enchytraeus albidus. Effect of Na-depletion on uptake rates of L-arginine from 10 μ M solutions after preincubation in appropriate incubation media for 4 h (15° C, 20 ‰ S, n = 5). n.s. indicates no significance ($p > 0.05$)

Composition of incubation medium	L-arginine	
	Uptake rate \pm s.e. (n moles/g/h), level of significance	Percentage
20 ‰ sea water	5.3 \pm 1.4	100
348 mM NaCl	4.0 \pm 0.3 n.s.	75
87 mM NaCl + 261 mM KCl	3.6 \pm 1.2 n.s.	68
87 mM NaCl + 261 mM LiCl	4.0 \pm 1.0 n.s.	75
87 mM NaCl + 261 mM choline-Cl	4.0 \pm 0.5 n.s.	75

When the oligochaetes were preincubated for 4 h in media containing 348 mM NaCl with a salinity equal to 20 ‰ sea-water controls (as measured by aerometry) or 87 mM (25 ‰) NaCl + 261 mM (75 ‰) KCl (LiCl, choline-Cl)—equimolar to 348 mM NaCl—a reduction of L-arginine absorption from 10 μ M solution by 25 ‰ became evident for 348 mM NaCl in comparison to 20 ‰ sea-water controls. Besides Na⁺, there seem to be one or more components in sea water, which are necessary for complete arginine transport. When 75 mole-% of the NaCl-solution were exchanged against K-, Li- or choline-Cl, no further reduction of arginine absorption could be realized. This is in contrast to neutral amino acid transport (glycine and L-valine) (SIEBERS & BULNHEIM, in press), which is regarded to be Na-dependent.

DISCUSSION

The above mentioned findings concerning differences in transport between neutral and basic amino acids provide sufficient information for several conclusions. Neutral amino acids are regarded to be actively transported against the concentration gradient from the medium across the body surface in *E. albidus*, as indicated by the following results, which are listed according to LEHNINGER (1974): (a) Strong inhibition of glycine and L-valine uptake by NaCN demonstrates dependence of the transport step on metabolic energy (SIEBERS & BULNHEIM, in press). (b) Uptake of amino acids is specific for a given substance. This specificity refers to the charge of amino acids and the position of the amino group (SIEBERS, in press) Neutral α -amino acids (glycine,

L-valine) are transported against the gradient in a saturable process, while basic amino acids (L-arginine and L-lysine) are obviously not concentrated in *E. albidus* and in *N. diversicolor* (Table 1). (c) The specific inward direction of neutral amino acid transport is demonstrated by SIEBERS & BULNHEIM (in press); in *E. albidus* glycine influx exceeded efflux by several times. (d) The uptake process of neutral amino acids (glycine, L-valine) is susceptible to selective inhibition by ouabain in *E. albidus*, demonstrating secondary active transport. Furthermore, neutral amino acid uptake in *E. albidus* depends on external Na^+ (SIEBERS & BULNHEIM, in press).

In contrast to neutral amino acids, uptake of basic amino acids (L-arginine, L-lysine) in *E. albidus* is extremely slow (Fig. 2), amounting to only few percentages of neutral amino acid absorption rates and not reaching equilibrium distribution ratios. For arginine the uptake curve is composed of a linear diffusion component and a non-linear saturable component (Fig. 1), which does not seem to be susceptible to metabolic inhibition by NaCN and ouabain and is therefore regarded to be independent of energy. Furthermore, arginine uptake is unmodified by depletion of ambient Na^+ . From the above mentioned findings on basic amino acid transport in *E. albidus* the saturable component of the uptake curve is regarded to fulfill the requirements for applying the term "facilitated diffusion", which means the energy-independent equilibration of a solute across a membrane, including saturation kinetics (Figs. 2, 3) and inhibition by molecules structurally analogous to the permeant considered.

For the gill of the common cockle, *Cerastoderma edule*, BAMFORD & MCCREA (1975) obtained evidence that L-lysine is absorbed by a saturable system differing from that for neutral amino acid uptake. Lysine uptake is also masked by diffusion at higher concentrations. The intestine of the polychaete *Arenicola marina* absorbs L-lysine against a concentration gradient (BAMFORD & STEWART, 1973). L-lysine is absorbed by the mid-gut of the bivalve *Mya arenaria* at a concentration ratio which differs only slightly from that for L-alanine and L-histidine (STEWART & BAMFORD, 1976).

For the body surface of the cestode *Hymenolepis diminuta* basic amino acids have not yet been demonstrated to be actively transported (PAPPAS & READ, 1975). However, the uptake system for basic amino acids in this cestode is clearly definable, since basic amino acids do not interact with neutral, acidic, or aromatic amino acids during absorption. These results are in contrast to the present findings for *E. albidus*. Also, in larvae of the cestode *Taenia crassiceps* a transport system is found which is specific for basic amino acids and not inhibited by neutral, dicarboxylic, and aromatic amino acids (HAYNES, 1970; PAPPAS & READ, 1973).

The arginine uptake system in the adult male trematode *Schistosoma mansoni* seems to resemble basic amino acid uptake in *E. albidus* more than that in cestodes, from which it differs considerably. Arginine uptake by *S. mansoni* was inhibited by neutral and basic amino acids, as was the case in methionine uptake (CHAPPELL 1974; PAPPAS & READ, 1975). As mentioned by PAPPAS & READ (1975), this is clearly different from cestodes, in which neutral and basic amino acids do not interact.

Na^+ -independent facilitated diffusion of amino acids was reported by WINTER & CHRISTENSEN (1964, 1965) for human and rabbit erythrocytes.

In vertebrate intestinal absorptive tissues a discrete transport mechanism for basic

amino acids exists, which is less efficient than that for neutral amino acids, operating at lower concentrations (WISEMAN, 1974). In sacs of everted small intestine of the rat, for instance, DL-ornithine and L-lysine are actively transported at a K_t -value and a v_{\max} of only about 10–20 % of neutral amino acid absorption data.

The preceding literature citations and present results do not provide sufficient information for conclusions concerning amino acid transport from a comparative view. On the other hand, the findings on amino-acid absorption in *E. albidus* confirm present concepts in regard to specificities of transport systems, which refer to the α -position of the amino group and the net charge of the amino acid molecule.

SUMMARY

1. Absorption of L-arginine across the body surface of the oligochaete *Enchytraeus albidus* and the polychaete *Nereis diversicolor* from 10 μM ambient concentrations proceeds at only few percentages of glycine and L-valine uptake rates. Distribution ratios do not exceed equilibrium. In *E. albidus* L-lysine is transported at the same low rates as observed for L-arginine.
2. In *E. albidus* the concentration dependent uptake of L-arginine consists of a saturable component and a diffusion component. The saturable component is susceptible to inhibition by structurally analogous substances. This inhibition is strongest in the presence of other basic amino acids and is still significant in the presence of neutral amino acids. Slight inhibition of L-arginine absorption is caused by L-glutamine and L-asparagine, while inhibition of arginine is only inconsiderable in the presence of acidic and β - and γ -amino acids. Arginine uptake rates are unchanged by organic acids and carbohydrates.
3. Because of nonsusceptibility of arginine transport to metabolic inhibitors (NaCN and ouabain) and Na^+ -depletion in *E. albidus*, the saturable component of arginine uptake is regarded to fulfill the requirements for applying the term "facilitated diffusion", which is discussed in relation to the active transport of neutral amino acids.
4. The mode of basic amino acid absorption is discussed in comparison to that in other animal species and absorbing tissues.

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