

Parasite structure of the Ocean Whitefish *Caulolatilus princeps* from Baja California, México (East Pacific)

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Abstract The metazoan parasite fauna of *Caulolatilus princeps* from northern Baja California, Mexico is quantitatively described for the first time. Further, the ecological aspects of prevalence, abundance, and intensity of infection are examined through an annual cycle. Six parasite species were recorded; 2 ectoparasites (1 monogenean and 1 copepod) and 4 endoparasites (2 digeneans and 2 nematodes). The digeneans *Choanodera caulolatlili* and *Bianium plicatum*, the nematodes *Anisakis* sp. and *Hysterothylacium* sp., and the copepod *Hatschekia* sp. set new geographical and host records. The highest values of prevalence and abundance were in *Anisakis* sp. (prevalence = 93.3%, abundance = 12.4 ± 4.7 ind/host) and in *Hysterothylacium* sp. (prevalence = 86.6%, abundance = 16.5 ± 3.4 ind/host). The mean intensity of infection showed maximum values in summer (August = 14.2) and minimums in winter (February = 4.2). The mean intensity was higher in *Hatschekia* sp. (20.3 ± 7.8) followed by *Hysterothylacium* sp. (18.6 ± 1.4) and *Anisakis* sp. (12.9 ± 2.2). Larval stages of *Anisakis* and *Hysterothylacium* were particularly important

due to their high abundance and prevalence, because they represent a human health risk (anisakiasis). In addition, the relationships between the metazoan parasites of *C. princeps* and host size and weight, fish condition and water temperature (bottom) are discussed.

Keywords Parasites · *Caulolatilus princeps* · Prevalence · Abundance · Intensity of infection · Mexico

Introduction

Knowledge about the parasite fauna of marine fishes from the Mexican Pacific coasts is still scarce (Pérez-Ponce de León et al. 1999; Sánchez-Ramírez and Vidal-Martínez 2002). The ocean whitefish, *Caulolatilus princeps* (Malacanthidae), ranges from Vancouver Island in British Columbia (Canada) to Peru, including the Galapagos Islands (Dooley 1978), and inhabits rocky reefs from 10 to 150 m depth (Hammann and Rosales-Casián 1990).

In the northwestern coasts of Mexico (both coasts of Baja California peninsula), this fish species is caught all year by coastal commercial and recreational fishing (Elorduy-Garay and Ruiz-Córdova 1998; Rosales-Casián and González-Camacho 2003; Siri-Chiesa and Moctezuma-Hernández 1989); the *C. princeps* catch during 2000 from Baja California Sur constituted 1,073 tons, 93% of the total catch for this species in Mexico (SAGARPA 2002). However, studies have focused on growth (Elorduy-Garay and Ramírez-Luna 1994), reproduction (Elorduy-Garay and Peláez-Mendoza 1996), and feeding (Elorduy-Garay et al. 2005), whereas the parasite community is poorly studied.

Therefore, the goals of the present study are (1) to identify the parasite fauna on *C. princeps* from the northwestern coasts of Baja California (México), (2) to determine the

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parasite community structure by their prevalence, abundance, intensity of infection and its variability through an annual cycle, and (3) to determine the relationships between parasite abundance and host size and weight, condition factor and water temperature.

Materials and methods

Ocean whitefish specimens were captured in the coast of San Quintín, Baja California, México by sport-fishing boats during 2005. The fishing area is in the Pacific Ocean outside of Bahía de San Quintín (30°33'37" N; 115°56'33" W), located 310 km south from the California border (USA) and 6 km from El Molino Viejo harbor (Old Mill). Fishing harvests pelagic and bottom fish species (Rosales-Casian and González-Camacho 2003; Rodríguez-Santiago and Rosales-Casián 2008) from surface to 150 m depth and as far as 50 km from rocky point Punta Entrada, outside of the bay. The whitefish individuals were captured from different rocky reefs in the area with random sampling which was dependent upon the boat catches.

Surface water temperatures (°C) during 2005 were obtained from boat captain reports. Temperatures at 50–140 m depth were obtained from station 107.32 of the IMECOCAL cruises (Lat. 30°27.288' N, Long. 116°9.696' W), located close to Isla San Martín (García-Córdova et al. 2005).

The site was sampled bimonthly, and parasite information was grouped by seasons: spring (April and June), summer (August), autumn (October), and winter (December and February). Ocean whitefish was identified using Miller and Lea (1972). All specimens were measured using total length (mm, LT) and weighed (g) with a digital balance Accu-Lab (6 kg); whitefish individuals ranged 370–510 mm LT, those lengths represented a range of 13–21 years old (Elorduy-Garay et al. 2005; Manríquez-Ledezma 2009). The Fulton's Condition Factor (Ricker 1975) was calculated for each specimen of *C. princeps* as: $K = [W/TL^3]$ 100,000 where: W = weight (g) and TL = total length (mm).

Specimens were dissected, and organs stored in plastic bags on ice. In the laboratory, internal organs (gills, liver, spleen, intestine, pyloric cecum, heart, gonads, and digestive tract) and external structures (skin and fins) were examined under a stereoscopic microscope, and all parasites were removed. Monogeneans and digeneans were fixed in AFA (acetic acid-formaldehyde-alcohol) solution for 2–24 h, then preserved in ethylic alcohol (70%), and stained with Gomori's trichromic stain (Vidal-Martínez et al. 2002). Nematodes were fixed in Berland's liquid, preserved in ethylic alcohol (70%), cleared with a solution of phenol-ethanol (Lent's solution), and mounted on slides

covered with glycerin-gelatin (Moravec et al. 1992). Copepods were first fixed in ethylic alcohol (70%), then cleared using a solution of glycerin-alcohol, and mounted on slides covered with glycerin-gelatin.

Identification of parasites was made using keys proposed by Yamaguti (1961, 1963, 1971), Vidal-Martínez et al. (2002), Bravo-Hollis (1967, 1982a, b) and Anderson et al. (1974–1983). To determine genera, Cressey and Boyle (1980, 1985), Kabata (1979, 1992a, b), and Boxshall (2004) were used. The parasitological material was deposited in the Laboratorio de Ecología Pesquera of the Centro de Investigación Científica y de Educación Superior de Ensenada, Baja California (CICESE), México.

Prevalence (%), abundance (number of parasites per host), and intensity (number of parasites/infected hosts) of parasites were determined according to Margolis et al. (1982). To assess significant variations in the mean abundance of parasites over seasons, a non-parametric analysis of variance of Kruskal-Wallis (KW) was performed (Steel and Torrie 1986). Spearman rank correlations were used to assess relationships between the abundance of parasites and host size and weight, fish condition and water temperature (bottom).

Results

Surface water temperature (°C) in the area showed a mean (\pm SE) of $16.0 \pm 0.3^\circ\text{C}$. The highest temperature mean was in August ($18.2 \pm 0.5^\circ\text{C}$), and lowest in February ($14.9 \pm 0.3^\circ\text{C}$). At fishing depth (50–140 m), annual temperature mean was $10.9 \pm 0.09^\circ\text{C}$ with highest in October ($11.5 \pm 0.20^\circ\text{C}$) and lowest in June ($10.2 \pm 0.08^\circ\text{C}$).

Species composition and organ specificity on *C. princeps*

A total of 91 specimens of *C. princeps* were examined (spring = 15 individuals; summer = 16; autumn = 38; and winter = 22). A total number of 3,820 parasites, belonging to 6 parasite species were identified (Table 1). They were 1 monogenean (*Choricotyle caulolatlili*), 2 digeneans (*Choanodera caulolatlili* and *Bianium plicitum*), 2 larval stages of nematodes (*Anisakis* sp. and *Hysterothylacium* sp.), and 1 copepod (*Hatschekia* sp.). The scientific names of *Choanodera caulolatlili* and *Choricotyle caulolatlili* are stated in full to avoid confusion.

The digestive tract was the most infested organ with 4 species. *Choanodera caulolatlili* and *B. plicitum* were found in intestine and stomach. *Choricotyle caulolatlili* and *Hatschekia* sp. were found on gills. *Anisakis* sp. was found in mesentery. Larvae of *Hysterothylacium* sp. were found in mesentery, stomach, intestine, and cecum (Table 1).

Table 1 Characterization of parasitic infections of *Caulolatilus princeps* from the coasts of San Quintin, Baja California, Mexico

Parasites	NF	PF	TNP	P	MA	MI	L
Monogenea							
<i>Choricotyle caulolatlili</i>	91	12	17	13.2	0.2 ± 0.1	2.6 ± 1.3	G
Digenea							
<i>Bianium plicitum</i>	91	57	247	62.6	2.7 ± 0.1	4.9 ± 1.1	I
<i>Choanodera caulolatlili</i>	91	34	108	37.3	1.3 ± 0.1	3.5 ± 0.9	I, S
Nematoda							
<i>Anisakis</i> sp.	91	84	1,113	92.3	12.4 ± 4.7	12.9 ± 2.2	M
<i>Hysterothylacium</i> sp.	91	78	1,462	85.7	16.5 ± 3.5	18.6 ± 1.4	M, S, I, IC
Copepoda							
<i>Hatschekia</i> sp.	91	52	873	57.1	10.1 ± 1.9	20.3 ± 7.8	G

Total number of fishes examined (NF), number of fishes parasitized (PF), total number of parasites per taxa (TNP), percentage of prevalence (P). Mean abundance (MA) and mean intensity (MI) of parasites (±SE) were calculated from mean values of seasons. Localization (L) in the host body; G Gills; IC Intestinal cecum; S Stomach; I Intestine; M Mesentery

Prevalence

The nematodes *Anisakis* sp. and *Hysterothylacium* sp. showed the highest prevalences (93.3 and 86.6%, respectively), and the monogenean *Choricotyle caulolatlili* the lowest (13.3%); the species *B. plicitum*, *Anisakis* sp., and *Hysterothylacium* sp. had prevalences higher than 60% and were considered as principal species; *Hatschekia* sp. and *Choanodera caulolatlili* were secondary species (58 and 38%, respectively), and *Choricotyle caulolatlili* with a prevalence of 13.3% was considered a satellite species (Table 1).

From late winter to autumn (February–October), *Anisakis* sp. showed a prevalence of 100% and a decrease (72%) in early winter (December) (Fig. 1a). Similarly, *Hysterothylacium* sp. exhibited prevalences of 100% from late winter to summer (from February to August), a slight decrease in autumn (95%, October), and an abrupt diminution in December (50%). The other species such as *Hatschekia* sp., *B. plicitum*, *Choanodera caulolatlili*, and *Choricotyle caulolatlili* showed an abrupt decrease in prevalence from spring to summer (Fig. 1a).

Abundance and intensity of parasites

The most abundant parasite species was *Hysterothylacium* sp. with a total of 1,462 individuals, and the lowest number was *Choricotyle caulolatlili* with 17 individuals (Table 1). The overall mean abundance (±SE) of parasites on *C. princeps* was 7.2 ± 0.7 ind/host, and seasonal mean abundances showed significant changes over time (Kruskall-Wallis, $H = 22.20$, $P = 0.0001$). *Hysterothylacium* sp. showed the highest overall mean abundance (calculated from average values of seasons) (16.5 ± 3.5 ind/host), followed by *Anisakis* sp. (12.4 ± 4.7 ind/host), *Hatschekia* sp. (10.1 ± 1.9 ind/host), *B. plicitum* (2.7 ± 0.1 ind/host), *Choanodera*

caulolatlili (1.3 ± 0.1 ind/host), and *Choricotyle caulolatlili* (0.2 ± 0.1 ind/host) (Fig. 1b).

The abundance of *Hysterothylacium* sp. was higher in late spring (21.6 ind/host) and lower in winter (7.8 ind/host), while *Anisakis* sp. showed the highest abundance in spring (16.4 ind/host) and lowest in winter (5.5 ind/host) (Fig. 1b). With respect to species, only the mean abundances of *Anisakis* sp. showed significant changes (Kruskall-Wallis, $H = 11.05$, $P = 0.011$) with the seasons.

With respect to the intensity of parasite infection, the highest mean value was exhibited by *Hatschekia* sp. (20.3 ± 7.8), followed by *Hysterothylacium* sp. (18.6 ± 1.4), and *Anisakis* sp. (12.9 ± 2.2); the rest of species had intensity values lower than 10 (Table 1). In the case of *Hatschekia* sp., the intensity increased from spring (14.6) to summer (40.2), which then decreased in winter (10.5). *Hysterothylacium* sp. showed the highest intensity in spring (21.6), followed by a decrease to the lowest (15.6) in winter (Fig. 1c). The rest of parasite species did not show important variations over time (Fig. 1c).

The Spearman rank correlations showed a significant positive correlation between prevalence and parasite abundance ($r = 0.943$, $P < 0.05$). Also, a significant correlation between the abundance of the nematode *Hysterothylacium* sp. and the surface water temperature ($r = 0.880$, $P < 0.05$) was detected. In addition, a significant negative correlation was found between the overall parasite abundance and the 50–140 m depth temperature ($r = -0.829$, $P < 0.05$). No correlations were found between the abundance of parasites with the host size and weight, and with fish condition.

Discussion

Four parasite species had been reported for ocean whitefish: 2 monogeneans: *Choricotyle caulolatlili* (as *Diclidophora*

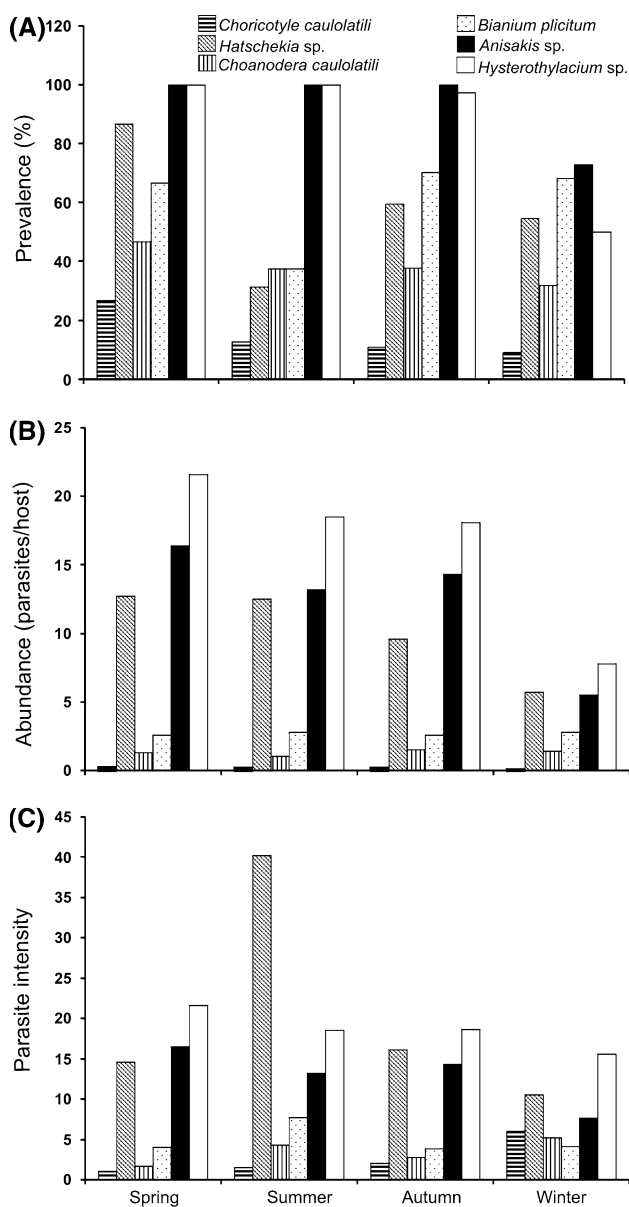


Fig. 1 a Prevalence (%), b abundance (parasites/host) and c intensity of parasites on specimens of *Caulolatilus princeps* collected at San Quintín, Baja California, during an annual cycle, 2005

caulolatlili, Meserve 1938) in Galapagos Islands (Merseve 1938) and *Jaliscia caballeroi* in Sonora Mexico (Bravo-Hollis 1982c), 1 digenean: *Proctoeces magnorus* in Isla Cedros Baja California, Mexico (Winter-Howard 1959), and 1 parasitic copepod: *Brachiella gracilis* in Ensenada, Baja California, México (Causey 1960). In our study, *Choanodera caulolatlili*, *B. plicitum*, *Hysterothylacium sp.*, *Choricotyle caulolatlili*, and *Hatschekia sp.* constitute new geographical records and *Anisakis sp.*, *B. plicitum*, *Hysterothylacium sp.*, and *Hatschekia sp.* were new host records.

The parasite composition of *C. princeps* was represented by 6 species, where digeneans contributed with 2 species

and 40% of the total individuals, this group of parasites are frequent in marine fishes (Rhode 1982; Castillo-Sánchez et al. 1998; Sánchez-Ramírez and Vidal-Martínez 2002; Muñoz et al. 2006).

The trematode species are common in marine and estuarine fishes of California, Oregon, and Washington (Love and Moser 1983). In the estuarine fish *Mugil cephalus*, a trematode frequency of 50% was found, and similar values in fishes from families Sciaenidae (*Leiostomus xanthurus* and *Micropogonias undulatus*), Scombridae (*Euthynnus lineatus*), and Khyphosidae (*Khyphosus elegans*) (Thoney 1993; León-Regagnon et al. 1997; Juárez-Arroyo and Salgado-Maldonado 1989). Parasite community structure may be influenced by factors such as the host biology, benthic habitat and territorial behavior (González and Poulin 2005).

The nematodes are important in fish parasitic diseases and are frequently found in different organs or microhabitats (Love and Moser, 1983; Alvarado-Villamar and Ruiz-Campos 1992; Thoney 1993; Castillo-Sánchez et al. 1998; Aloo et al. 2004). In the present study, parasites with high specificity were the ectoparasites *Choricotyle caulolatlili* and *Hatschekia sp.*, which were found on gills only; *Anisakis sp.* and *B. plicitum* were found in the mesentery and intestines, respectively. This specificity could be due to the kind of nutrients that these organs offer.

Larvae of *Hysterothylacium sp.* and *Anisakis sp.* were the most abundant parasites in the ocean whitefish and accounted for 67.4% of total parasites. Most parasite groups, which infect marine fishes, are at adult stages, indicating that fishes are important as final hosts rather than as intermediate hosts (Juárez-Arroyo and Salgado-Maldonado 1989; Castillo-Sánchez et al. 1998; León-Regagnon et al. 1997). Previous studies have suggested that some parasites of *C. princeps* come from larvae hosted in intermediate hosts such (mollusk gastropods, cephalopods, fishes), which harbor mainly digeneans whose abundance depends on diet (Elorduy-Garay et al. 2005). Furthermore, demersal fish studies indicated that many species are intermediate links in the marine food chain (Muñoz et al. 2006; Oliva and Luque 1998; Cordeiro and Luque 2004; Sánchez-Ramírez and Vidal-Martínez 2002).

In this study, larvae of *Hysterothylacium sp.* showed higher values of prevalence (87%) and parasite intensity (18.5 parasites/fish infected) than those reported for *Hysterothylacium aduncum* in Chilean salmon farms (prevalence = 79.1%; mean intensity = 4.9 parasites/fish infected) (González 1998). A common pattern in marine fish is that parasite intensity shows variability by species complexity with low prevalences and abundances (Valtonen et al. 2001; Tavares and Luque 2004).

Our study indicated that parasite abundance was not correlated with size of *C. princeps*; however, we did not analyze smaller sizes that are unavailable for sport-fishing.

Nevertheless, Poulin (2000) documented a significant correlation between fish length and intensity of infection for cestodes, larval digeneans, and gnathiid isopods. The condition of *C. princeps* was also independent of number of parasites hosted. This independence was similar to brown trout (*Salmo trutta*) from Fernworthy, Devon, United Kingdom (Kennedy and Lie 1976), where the parasites did not alter the condition.

Finally, larvae of *Hysterothylacium* sp. and *Anisakis* sp. were relatively abundant in the ocean whitefish and were important for the anthropocenoses that can develop. These parasite species can be infective to humans and cause Anisakiasis, and fish which have been infected with *Anisakis* spp. can produce an anaphylactic reaction in people who have become sensitized to immunoglobulin E (Domínguez-Ortega et al. 2001). Therefore, we recommend not eating raw or inadequately cooked fish. The present study is the first parasitological record for this fish species in the north-western coasts of Baja California. Further, our results on the prevalence, abundance, and intensity of parasites over an annual cycle helped us to diagnose the health status of this important marine fish for the region.

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References

- Aloo PA, Anam RO, Mwangi JN (2004) Metazoan parasites of some commercially important fish along the Kenyan coast. *West Indian Ocean J Mar Sci* 3:71–78
- Alvarado-Villamar MR, Ruiz-Campos G (1992) Estudio comparativo del grado de infección de macroparásitos en seis especies de *Sebastes* (Pisces Scorpenidae) de la costa Noroccidental de Baja California, México. *Cien Mar* 18:79–92
- Anderson RC, Chabaud AG, Willmott S (eds) (1974–1983) CIH keys to the nematode parasites of vertebrates, vol 1–10. Commonwealth Agricultural Bureaux, Farnham Royal, Bucks: England
- Boxshall GA (2004) An introduction to copepod diversity. Ray Society Series 166. With SH. Halsey. 2 Parts, vols. The Ray Society of London, United Kingdom, p 940
- Bravo-Hollis M (1967) Helmintos del Pacífico mexicano XXV. Descripción de tres monogéneos del Golfo de California. *Anal Inst Biol Univ Nac Aut Méx Ser Zool* 37:107–123
- Bravo-Hollis M (1982a) Helmintos de peces del Pacífico mexicano XXXVII. Sobre seis especies conocidas de monogéneos del Suborden Microcotylina Lebedev, 1972. *Anal Inst Biol Univ Nac Aut Méx Ser Zool* 52:1–12
- Bravo-Hollis M (1982b) Helmintos de peces del Pacífico mexicano XXXIX. Estudio de Monogéneos del suborden Microcotylina Lebedev, 1972, con la presentación de una subfamilia y una especie nuevas. *Anal Inst Biol Univ Nac Aut Méx Ser Zool* 53:13–26
- Bravo-Hollis M (1982c) Helmintos de peces del pacífico Mexicano. XXXIX. Dos subfamilias nuevas de Monogéneos de la familia Macrovalvitrematidae Yamaguti, 1963. *Anal Inst Biol Univ Nac Aut Mex Ser Zool* 52:27–38
- Castillo-Sánchez E, Rosales-Casián J, Pérez-Ponce de León G (1998) Helminth parasites of *Paralichthys californicus* (Osteichthyes: Paralichthyidae) in Estero de Punta Banda, Todos Santos Bay and San Quintín Bay, Baja California, México. *Cien Mar* 24:443–462
- Causey D (1960) Parasitic Copepoda from Mexican coastal fishes. *Bull Mar Sci Gulf Caribb* 10:323–337
- Cordeiro AS, Luque JL (2004) Community ecology of the metazoan parasites of Atlantic moonfish, *Selene setapimmis* (Osteichthyes: Carangidae) from the costal zone of the State of Rio de Janeiro, Brazil. *Braz J Biol* 64:399–406
- Cressey R, Boyle-Cressey H (1980) Parasitic copepods of Mackerel and tuna-like fishes (Scombridae) of the world. *Smith Contrib Zool* 311:54–61
- Cressey R, Boyle-Cressey H (1985) *Holobomolochus* (Copepoda: Bomolochidae) redefined, with descriptions of three new species from the eastern pacific. *J Crust Biol* 5:717–727
- Domínguez-Ortega J, Alonso-Llamazares A, Rodríguez L, Chamorro M, Robledo T, Bartolomé JM, Martínez-Cócera C (2001) Anaphylaxis due to hypersensitivity to *Anisakis simplex*. *Int Arch Allergy Immunol* 125:86–88
- Dooley JK (1978) Systematics and biology of the tilefishes (Periformes: Branchiostegidae and Malacanthidae) with description of two new species. NOAA technical report. NMFS Circular 411:25–27
- Elorduy-Garay JF, Peláez-Mendoza AK (1996) Hábitos alimenticios de *Caulolatilus affinis*. Gill 1865 (Perciformes: Branchiostegidae) en la Bahía de La Paz, B.C.S., México. *Rev Biol Trop* 44:241–249
- Elorduy-Garay JF, Ramírez-Luna S (1994) Gonadal development and spawning of the ocean whitefish, *Caulolatilus princeps* Jenys 1842 (Pises: Branchiostegidae) in the Bay of La Paz, B.C.S., México. *J Fish Biol* 44:553–566
- Elorduy-Garay JF, Ruiz-Córdova SS (1998) Age, growth, and mortality of *Caulolatilus affinis* (Osteichthyes: Branchiostegidae) from the Southern Gulf of California. *Pac Sci* 52:259–272
- Elorduy-Garay JF, Ruiz-Córdova SS, Díaz JG (2005) Age, growth and mortality of *Caulolatilus princeps* (Pisces: Malacanthidae) from the southern Gulf of California. *Hidrobiol* 15:289–297
- García-Córdova J, Robles-Pacheco JM, Gómez-Valdés J (2005) Informe de datos de CTD. Campaña IMECOCAL 0501/02. B/O Francisco de Ulloa. Enero 21-Febrero 10 de 2005. Informe Técnico, Departamento de Oceanografía Física CICESE, pp 132
- González L (1998) The life cycle of *Hysterothylacium aduncum* (Nematoda: Anisakidae) in Chilean marine farms. *Aquaculture* 162:173–186
- González MT, Poulin R (2005) Spatial and temporal predictability of the parasite community structure of a benthic marine fish along its distributional range. *Int J Parasitol* 35:1369–1377
- Hamman GM, Rosales-Casián JA (1990) Taxonomía y estructura de la comunidad de peces del Estero de Punta Banda y Bahía de Todos Santos, B.C., México. In: Rosa-Vélez J, González-Farías (eds) Temas de Oceanografía Biológica en México. Universidad Autónoma de Baja California, Ensenada, p 337
- Juárez-Arroyo AJ, Salgado-Maldonado G (1989) Helmintos de la "lisa" *Mugil cephalus* Lin en Topolobampo, Sinaloa, México. *Anales del Instituto de Biología de la Universidad Nacional Autónoma de México* 60:279–298
- Kabata Z (1979) Parasitic copepoda of British fishes. Ray Society, London, p 468

- Kabata Z (1992a) Copepods parasitic on fishes. Synopses of the British Fauna (new series) No. 47. Published for The Linnean Society of London and The Estuarine Coast Sci Assoc, Great Britain, p 264
- Kabata Z (1992b) Copepoda parasitic on Australian fishes, XV. Family Ergasilidae (Poecilostomatoida). *J Nat Hist Lond* 26:47–66
- Kennedy CR, Lie SF (1976) The distribution and pathogenicity of larvae of Eustrongylides (Nematoda) in brown trout *Salmo trutta* L in Fernworthy Reservoir Devon. *J Fish Biol* 8:293–302
- León-Regagnon V, Pérez-Ponce de León G, García-Prieto L (1997) Description of *Heteroplectanum oliveri* sp n. (Monogenea: Diplectanidae) and comments on the helminth fauna of *Kyphosus elegans* (Perciformes: Khyphosidae) from Chamela Bay, México. *J Hel Soc of Wash* 64:9–16
- Love MS, Moser M (1983) A checklist of parasites of California, Oregon and Washington marine and estuarine fishes. US Dept. Commer., NOAA Tech. Rep., NMFS SSRF-777, pp 576
- Manríquez-Ledezma Y (2009) Edad y crecimiento de *Caulolatilus princeps* (Jenyns, 1840) en Bahía de La Paz y Bahía Magdalena, B.C.S. México. M.Sc. Thesis, CICIMAR-IPN, La Paz BCS, México, pp 56
- Margolis LG, Esch JC, Holmes AM, Kuris SchadGA (1982) The use of ecological terms in parasitology (report of an ad hoc committee of the American Society of Parasitologists). *J Parasitol* 68:131–133
- Merseve FG (1938) Some monogenetic trematodes from the Galapagos Islands and the neigh-boring Pacific. *Univ South Calif Allan Hancock Found Publ Occas Pap* 2:30–89
- Miller DJ, Lea RN (1972) Guide to the coastal marine fishes of California. California Dept Fish and Game Fish Bull 157:1–249
- Moravec FV, Nasincord, Scholz T (1992) Methods of investigation of endoparasitic helminths, Training of fish Parasites, Institute of Parasitology, Acad Sci Ceske Budejovice, p 54
- Muñoz G, Grutter AS, Cribb TH (2006) Endoparasite communities of five fish species (Labridae: Cheilininea) from Lizard island: how important is the ecology and phylogeny of the host? *Parasitology* 132:363–374
- Oliva ME, Luque JL (1998) Metazoan Parasite Infracommunities in five Sciaenids from the Central Peruvian coast. *Mem Inst Oswaldo Cruz* 93:175–180
- Pérez-Ponce de León G, García-Prieto L, Mendoza-Garfias B, León-Regagnon V, Pulido-Flores G, Aranda-Cruz CV, García-Vargas F (1999) Listados faunísticos de México. IX. Biodiversidad de helmintos parásitos de peces marinos y estuarinos de la Bahía de Chamela, Jalisco. Universidad Autónoma de México, México, p 51
- Poulin R (2000) Variation in the intraspecific relationship between fish length and intensity of parasitic infection: biological and statistical causes. *J Fish Biol* 56:123–137
- Rhode K (1982) Ecology of marine parasites. University of Queensland Press. St. Lucia, Queensland, p 245
- Ricker WE (1975) Computation and interpretation of biological statistics of fish populations. *Bull Fish Res Bd Can* 191:1–382
- Rodríguez-Santiago MA, Rosales-Casián JA (2008) Abundance and size composition of vermilion rockfish, *Sebastes miniatus* (Jordan and Gilbert 1880), from sport fishing catches of San Quintín, Ensenada, Baja California, México. *Bull South Cal Acad Sci* 107:25–32
- Rosales-Casian JA, Gonzáles-Camacho JR (2003) Abundance and importance of Fish Species from the Artisanal Fishery on the Pacific Coast of Northern Baja California. *Bull South Cal Acad Sci* 102:51–65
- SAGARPA (2002) Anuario Estadístico de Pesca 2000, Secretaría de Agricultura, Ganadería, Desarrollo Rural. Pesca y Alimentación, México, p 268
- Sánchez-Ramírez C, Vidal-Martínez V (2002) Metazoan parasite infracommunities of Florida pompano (*Trachinotus carolinus*) from the coast of the Yucatán Peninsula, México. *J Parasitol* 8:1087–1094
- Siri-Chiesa M, Moctezuma-Hernández P (1989) La pesca de Baja California. Universidad Autónoma de Baja California: México, 211
- Steel RG, Torrie JH (1986) Bioestadística: principios y procedimientos. McGraw-Hill, México, p 662
- Tavares LER, Luque JL (2004) Community ecology of the metazoan sea catfish, *Netuma barba* (Osteichthyes: Ariidae), from the coastal zone of the State of Rio de Janeiro, Brazil. *Braz J Biol* 64:169–176
- Thoney D (1993) Community ecology of the parasites of adult spot, *Leiostomus xanthurus* and Atlantic croaker, *Micropogonias undulatus* (Scianidae) in the Cape Hatteras region. *J Fish Biol* 43:781–804
- Valtonen ET, Pulkkinen K, Poulin R, Julkunen M (2001) The structure of parasite component communities in brackish water fishes of the north-eastern Baltic Sea. *Parasitology* 112:471–481
- Vidal-Martínez V, Aguirre-Macedo L, Sholz T, González-Solis D, Mendoza-Franco F (2002) Atlas de los helmintos parásitos de cíclidos de México. Instituto Politécnico Nacional, México, p 183
- Winter-Howard A (1959) Some trematodes from fishes of the Mexican and adjoining California Pacific, with notes on the distribution of eastern Pacific fish trematodes. Ph. D. Thesis, University of Southern California, p 198
- Yamaguti S (1961) Systema Helminthum. III. Nematodes. Interscience Publishers, INC, New York, p 1261
- Yamaguti S (1963) Systema Helminthum. Vol. IV: Monogenea and Aspidocotylea. Interscience Publishers, New York, p 381
- Yamaguti S (1971) Synopsis of digenetic trematodes of vertebrates, vol I. Keigaku Publishing Co, Tokio Japan, p 1074